Massachusetts Water Resources Authority | Pathogen Monitoring Program | 2008 Review 617-242-6000 – www.mwra.com

Samples at Cosgrove and CVA Intakes

Even though testing for *Giardia* and *Cryptosporidium* was not required by EPA until 2006, MWRA has been monitoring for them in source waters since 1994. EPA does set inactivation requirements for *Giardia*, while requirements for *Cryptosporidium* will not come into effect until 2014. MWRA provides disinfection for both *Giardia* and *Cryptosporidium*. *Giardia* and *Cryptosporidium* sampling was initiated in 1994 (monthly samples at Shaft 4, later moved to Cosgrove Intake). In mid-January 1999, sampling was increased to weekly at Cosgrove Intake. Starting on April 1, 2008, sampling was moved to Carroll Water Treatment Plant Inlet. Biweekly sampling is conducted at the CVA Intake.

Giardia and Cryptosporidium results are reported as number of cysts per 100 L. Until March 2004, MWRA used the EPA-approved method, ASTM D19 (ICR) with 100 L samples. Under this method, identifications were grouped into 2 categories: presumed (no internal structures identified) and confirmed (one or more internal structures identified). From July 1997 to March 2004, no samples confirmed positive for Giardia, and no samples were presumptive or confirmed positive for Cryptosporidium. In April 2004, MWRA began testing samples using the newly approved EPA Method 1623 with 50 L samples, which is about twice as sensitive as the ICR method (3 to 4 times the recovery rate with half the volume.) Under the new method, identifications are grouped into 3 categories: empty (no internal structures), amorphous structure (structure not consistent with a normal organism), and one or more internal structures. The results for 2008 using Method 1623 are listed below.

Wachusett: Cryptosporidium Results for MetroBoston: Jan 2008 - Dec 2008

53 Samples Collected

Number of Positive Samples	Total Number of Oocysts Detected	Empty Oocysts	# with Amorphous Structure	# with One or More Internal Structures	Average (oocysts/100L)	Range of Detects (oocysts/100L)
1	1	1	0	0	0.037/100L	1

Wachusett: Giardia Results for MetroBoston: Jan 2008 – Dec 2008

53 Samples Collected

Number of Positive Samples	Total Number of Cysts Detected	Empty Cysts	# with Amorphous Structure	# with One or More Internal Structures	Average (cysts/100L)	Range of Detects (cysts/100L)
23	48	44	3	1	1.83/100L	1 - 11

CVA Intake: Cryptosporidium Results for Chicopee Valley Aqueduct: Jan 2008 – Dec 2008

26 Samples Collected

Number of	Total Number of	Empty	# with	# with One or More	Average	Range of
Positive	Oocysts Detected	Oocysts	Amorphous	Internal Structures	(oocysts/100L)	Detects
Samples			Structure			(oocysts/100L)
2	2	2	0	0	0.15/100L	1

CVA Intake: Giardia Results for Chicopee Valley Aqueduct: Jan 2008 – Dec 2008

26 Samples Collected

20 Sumples Conceicu								
Number of	Total Number of	Empty	# with	# with One or More	Average	Range of Detects		
Positive	Cysts Detected	Cysts	Amorphous	Internal Structures	(oocysts/100L)	(cysts/100L)		
Samples			Structure					
2	4	4	0	0	0.31/100L	1 - 3		

Note: A complete record of results can be found on the MWRA website at www.mwra.com.

Research Efforts

MWRA is currently engaged in a voluntary, joint research effort with Tufts University looking at levels of *Cryptosporidium* and *Giardia* in drinking water using a high volume sample (1000 liters). This monitoring is part of a larger multi-city study looking at levels of *Cryptosporidium* exposure in the population and potentially related levels in drinking and recreational waters. Since the routine, EPA-approved ICR method previously used by the MWRA had few detects, no statistical comparisons of human exposure to drinking water were possible. As a result, MWRA and Tufts decided to use a more sensitive method to determine the variability, if any, of levels of *Cryptosporidium* and *Giardia*.

The research monitoring started using a weekly composite sample (some water each day for the entire week) of 1,000 liters at Shaft 9A, a site within the water system that is representative of water delivered to customers in the MetroBoston system. In 2008, bi-weekly composite samples of 2,000 liters were used. The water is then evaluated using a test method basically the same as Method 1623. All *Cryptosporidium* oocysts and *Giardia* cysts, both confirmed and empty, are counted. This method, using a large sample volume, is more than 20 times more sensitive than the present sampling protocol with Method 1623 now used by MWRA, and at least 40 times as sensitive as the previous ICR method.

The data collected so far is generally consistent with MWRA's past data. As was expected, the much higher sample volumes and the more sensitive testing have yielded some positive samples; all but one of the positives has been below the nominal detection limit of Method 1623 (1-oocyst/50 liters), and the overall average for all samples since 2001 is 0.033 oocysts/100 liters. Tufts has also tested for *Giardia* using the same testing method as above, and the overall average is 0.022 cysts/100 liters since 2002.

Research Sampling - Cryptosporidium Results: Feb 28, 2008 - Dec 26, 2008

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Number of	Total Number	Empty Oocysts	# with	# with One or	Average	Range of
Positive	of Oocysts		Amorphous	More Internal	(oocysts/100L)	Detects
Samples	Detected		Structure	Structures		(oocysts/100L)
3	3	3	0	0	0.008/100L	0.05

Research Sampling - Giardia Results: Feb 28, 2008 – Dec 26, 2008

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Number of	Total Number	Empty Cysts	# with	# with One or	Average	Range of
Positive	of Cysts		Amorphous	More Internal	(cysts/100L)	Detects
Samples	Detected		Structure	Structures		(cysts/100L)
1	1	0	1	0	0.003/100L	0.05

Testing Limitations and Response Protocol

It is important to note that *Cryptosporidium* and *Giardia* monitoring has significant limitations. The tests do not clearly distinguish between live and dead cysts, cannot determine if an organism is in fact infectious to humans, and the infectious dose of various strains of *Cryptosporidium* is not well understood. Nonetheless, in 1996, MWRA adopted a trigger level of 10 oocysts per 100 liters (recommended by Rose and Haas, leading researchers in pathogen and risk/health analysis) above which notification and other actions would be undertaken. Total number of positives, both confirmed and empty, are included in this trigger level. No special actions are required for levels below this level. Even with the new, more sensitive testing method, the average level found is well below the 10-oocyst per 100 liter trigger level. Furthermore, MWRA's new treatment with ozone at the Carroll Treatment Plant is capable of inactivating (killing) at least 99.9% of any *Giardia* and at least 99% of any *Cryptosporidium* which may be present and viable.

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¹ The new 2008 contract with Tufts was not finalized until late February, so there were only 20 samples for the year.